

DESINFECTEST®

The latest generation of prepared dip slides for industrial, food and environmental microbiological testing. These kits are easy-to-use, enabling them to be used by non-specialised staff, obtaining highly reliable results. They can be used for two types of testing:

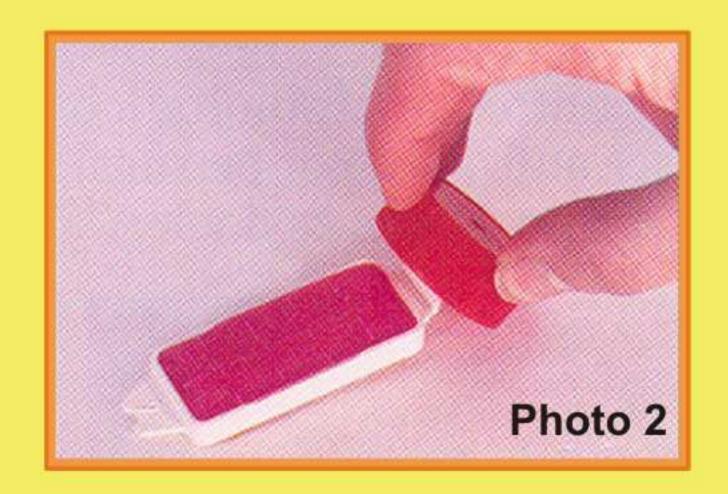


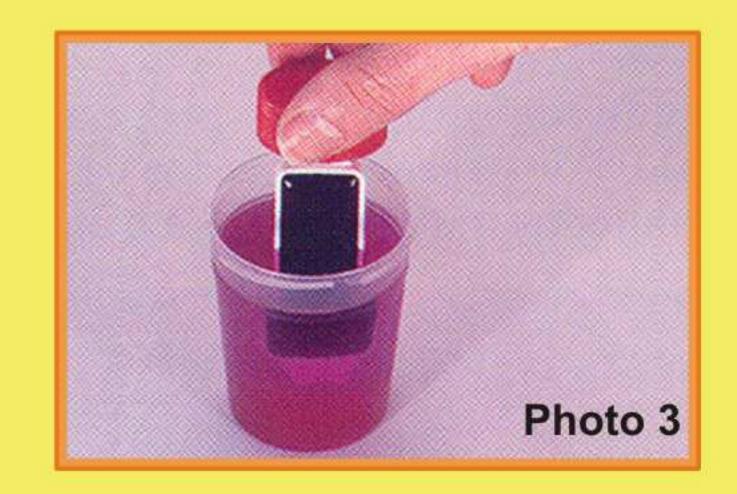
- 1. For the **quantitative testing of surfaces**, by contact. In HACCP, it facilitates the detection and testing of critical points. The flexible slide makes full contact easier between the culture medium and the surface to be analysed. Sensitivity is far higher than that of the swab wiping method. It is more convenient than using contact plates.
- 2. For the **semi-quantitative testing of raw liquids**, by immersion. The advantage is that it saves handling the endless dilutions of the traditional method, with optimal ease of use. Applicable in samples with usual contamination of over 10³ ufc/ml.

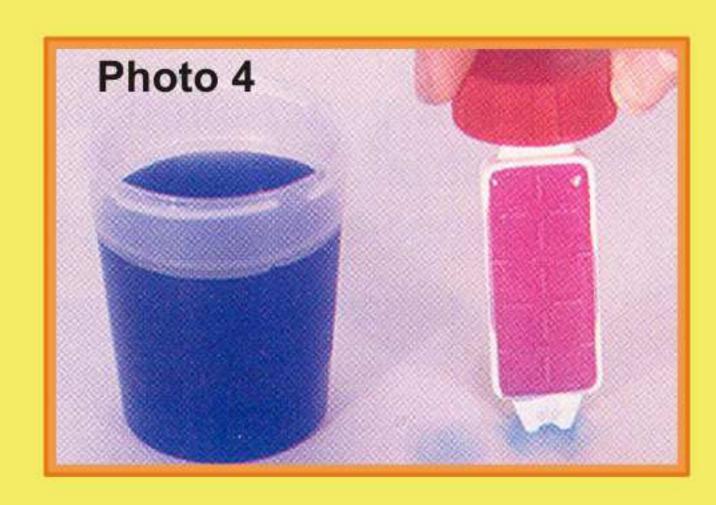
In both cases, the sealed slide format makes them easier and safer to handle and transport. A complete range is available for the analysis of all microbial indicators (photo 1):

- Total count after disinfection TSA Neutralizing cream -TCC/PCA-TCC: D-AEM/AET, MBN101
- Environmental yeasts and moulds (pink) + Dermatophyte fungi (orange): D-LM/DTM, MBN102
- Enterobacteria, including Salmonella (green) + Coliforms, including E.coli (cream): D-EC, MBN103
- Staphylococcus aureus (cream) + Listeria including L.monocytogenes (red): D-STAPH/LIS, MBN106
- DESINFECTEST®-MIX (colourless face: Bacteria / pink face: Fungi), MBN407
- DESINFECTEST®-ARICPC (colourless face: Bacteria / purple face: Enterobacteriaceae), MBN200

Store away from light at 15-25°C, avoiding sudden changes in temperature. Expiry date is 6 months after date of manufacture. Only open immediately prior to use. Do not touch the media. Do not reuse. Condensation water is normal if it does not succeed 1 ml. DO NOT use if the media are discoloured, contaminated or very dry. Autoclave or fill with bleach after use.











Surfaces:

Unscrew and apply for 10 seconds to the surface, without rubbing, applying light pressure (photo 2), preferably putting a finger of the other hand on the end of the plastic strip. The flexibility of the slide ensures full contact with the surface. Repeat these steps for the other face on a nearly surface. The most reliable sample is of 100 cm² (5 slides on both sides), due to the contagious distribution of micro-organisms on surfaces. A larger sample is a waste, but a smaller sample is not representative enough for low-contaminated surfaces.

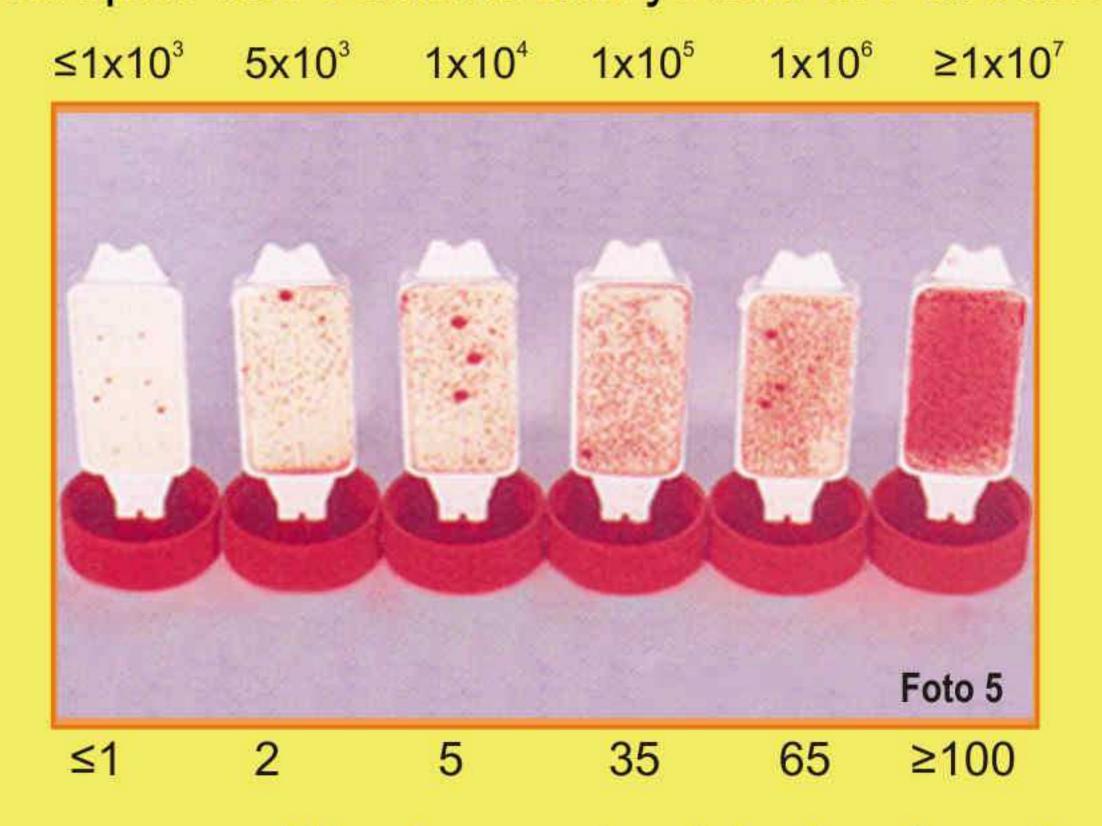
Líquids:

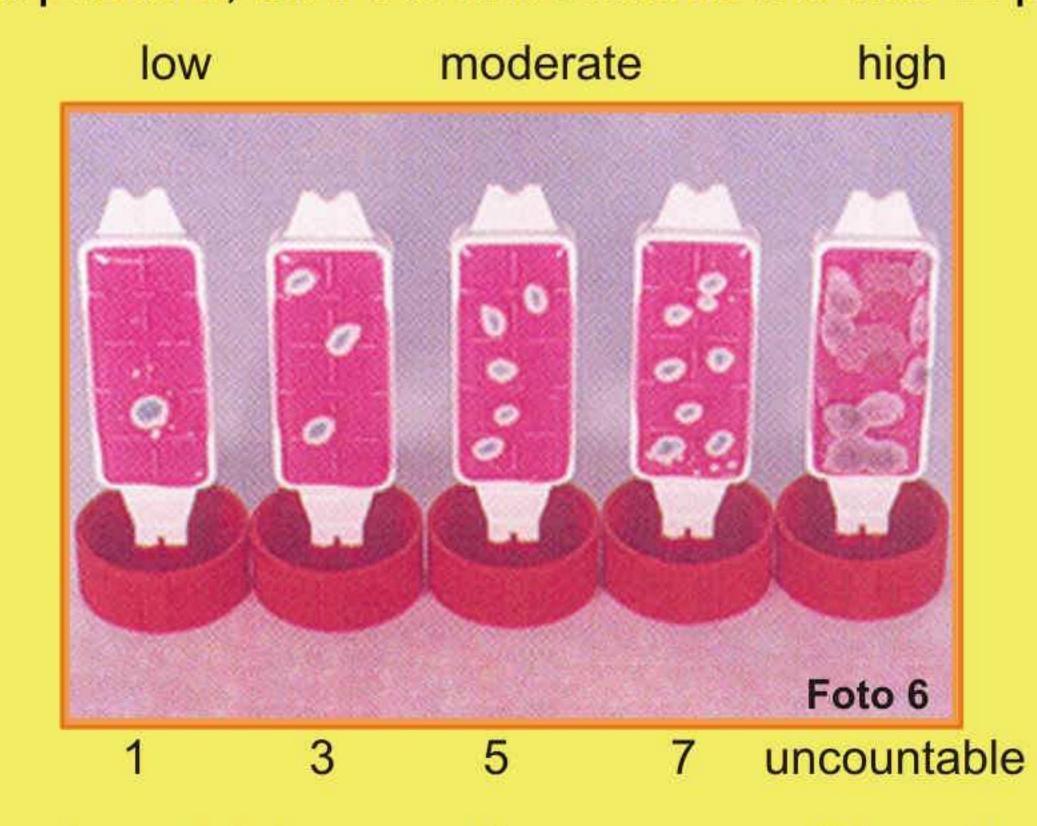
Unscrew and immerse in the liquid for a few seconds (photo 3). If the liquid is rather viscous or is expected to be highly contaminated, dilute it with sterile dilutant, such as 1 ml of sample in 100 ml of RINGER and then multiply the results by their dilution factor, in this case 100. Never shake, but simply allow the surplus liquid to drip, resting the end of the strip on absorbent paper (photo 4).

In both cases, replace each slide inside its tube. Incubate in a vertical position (preferably with the cap upwards and not fully tightened), 1 – 3 days at 21-37°C (economical MICROKIT incubator, VMT051). The chosen temperature depends on if you are looking for pathogenic human flora (35-37°C) or spoiling saprophyte flora (21-25°C, or the normal temperature of the sample). These are two different microbial populations, the count or presence of one is not comparable to the other (independently of whether they are bacteria or fungi). The best method is to test for both populations, performing duplicate DESINFECTEST® at both temperatures.

INTERPRETATION OF RESULTS:

If you do not wish to count, you can use the adjoining table to help with enumeration. The upper numbers are semi-quantitative and are ufc/ml of liquid sample. The lower numbers are quantitative and are ufc/cm² of surface sample. The bacteria and yeasts are shown in photo 5, and the filamentous moulds in photo 6.





Establish your own self-testing scales (starting from the value which normally causes problems) and test routinely to for prevention using DESINFECTEST ®. As a guide, for surfaces there should not be (according to APHA and various authors) more than 4 ufc/100 cm² on clean instruments and areas, or more than 60 ufc/100 cm² on tables, in restaurants, nurseries, butcher's, washbasins, etc. or more than 200 ufc/100cm² in abattoirs, on floors or in lavatories. According to UNE 100012, there should no more than 4 ufc/cm² on surfaces prior to disinfection. Furthermore, the absence or minimisation of pathogens (*E.coli, S.aureus*, Salmonella, *L.monocytogenes*) is compulsory. Warning: loss of shine in the medium may be a sign of counts so high that the human eye cannot detect them (thousands of tiny colonies). In this case, repeat after diluting. Important: if the medium turns to the colour that is characteristic of the colonies, as defined below, it is a sign of extremely high contamination.

Bacteria (including Bacillus, Pseudomonas, Enterococcus and Micrococcus) grow in TSA-Neutralizing + TTC/PCA+TTC, reddish (red medium= huge count). Moulds (including Aspergillus, Alternaria, Mucor and Penicillium) grow in Rose Bengal Agar, pink, as filaments and yeasts (including Candida, Pichia, Saccharomyces and Yarrowia) as dots. No bacteria grow in this medium, due to the Chloramphenicol. Dermophyte fungi (such as Trichophyton, Epidermophyton and Microsporum) grow in DTM, orange, as filaments that turn the medium red. No bacteria grow in this medium either. Enterobacteria grow in Hektoen, green, with various colours; Salmonella with blue-black colonies; E.coli with salmon-coloured colonies (salmon or black medium= huge count). Coliforms grow in Mug Plus Agar, with pink colonies and *E.coli*, with blue colonies. Total coliforms are pink + blue colonies. *S.aureus* grows in Baird Parker, cream, with black colonies surrounded by a colourless halo. *L.monocytogenes* grow in Palcam, red, with grey colonies surrounded by a grey/brown -black halo (black media indicate huge counts).