

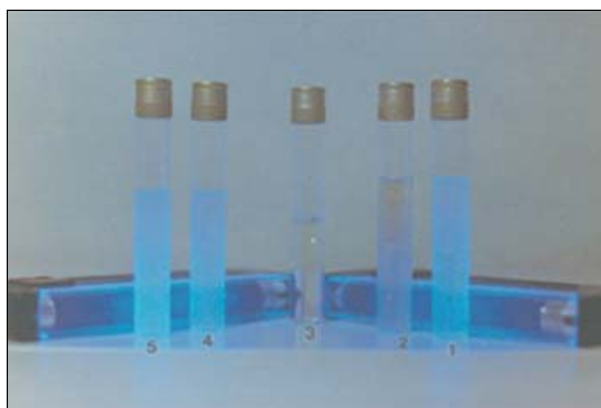
MCC-COLICULT ChromoFluorogenic Broth

Detection Presence/Absence of Coliformes and E.Coli (included E.Coli O157) in water, also in food.

COMPOSITION

Tryptose	20,00 g
Hidrogenphosphate di-K	2,75 g
K dihidrogenphosphate	2,75 g
Chloride sodic	5,00 g
Lactose	5,00 g
Laurilsulphate sodic	0,10 g
Tryptophane	1,00 g
MUG	0,10 g
Chromogen (X-Gal)	c.s.

(Formula per liter)
 pH final: 6,8 ± 0,1



MCC-COLICULT is Lauryl MUG Broth with X-Gal
 1, 4, 5: *E.coli* positive (fluorescent)
 2, 3: *E.coli* negative (no fluorescent)

PREPARATION

Add 1 g sterilized by irradiation (VIALS FPA900) to 100 ml of sample water, which will do as solvent; don't sterilize.

Or, add 10 g/l of final medium not irradiated (DMT900-) and autoclave 5 minutes to 116°C, inoculating after.

FOR EXCLUSIVE USE OF LABORATORY.

KEEP BOTTLE CLOSED IN A DRY, FRESH AND DARK PLACE.

DEHIDRATED CODE: [DMT900-](#)

QUALITY CONTROL OF MEDIUM

Elaborated in our laboratory; it is prudent repeat in your laboratory always conditions change (more than three months without use, after disinfect laboratory, after keep to higher temperature, when it takes bizard aspects althoug expire date is correct,...)

Shake bottles P/A (RPL303) before use it, to ensure homogenize of temporary gradients of density of components: crystallised/precipitated can be dissolved when you add 100 ml of sample.

DEHIDRATED: beige powder, sterile.

PREPARED: sterile, lightly straw.

QUANTITATIVE GROWING CONTROL 18-24 h at 35-44'5 °C more or less:

Escherichia coli MKTA 25922: Turns to blue in 18 hours. Fluorescence intense under UV light of 366 nm. Indol Kovacks + (red in surface)

Escherichia coli MKTA 35150 (O157:H7): Turns to blue in 18 hours. Without fluorescence under UV light of 366 nm. Indol Kovacks + (red in surface)

Enterobacter aerogenes MKTA 13048: Turn to turquoise blue in 18 hours. Without fluorescence in light of 366 nm. Indol kovacks – (not red in surface)

Pseudomonas aeruginosa MKTA 10145: Growing without turning neither fluorescence

Enterococcus faecalis MKTA 29212: Inhibited.

PRESENTATION: BOTTLES P/A 18 ml (RPL303), TUBES 10 ml (TPL637), VIALS PREWEIGHTED AND IRRADIATED 1 g (FPA900), DEHIDRATED 100 g (DMT900-)

INSTRUCTIONS AND RESULTS INTERPRETATION

-Add 1 g of irradiated medium (FPA900) to 100 ml of water (or 2 g to 250 ml of bottling water), for detection of Presence/Absence of Coliforms and E.Coli. Unic caution is operator don't touch neither water nor powder with their hands, to avoid artificial contaminations. It is necessary treat previously chlorated water, by tiosulphate sodic, so damaged cells will be recovered.

-If you use prepared bottles P/A (RPL303), add 100 ml of sample water to the bottle, which contain inactivators for chlorine.

-If you use prepared tubes (TPL637), you just add 1 ml of sample or food dilution.

-If you use dehydrated medium no-sterile (DMT900-), you have to add 10 g to 1 liter of bidestiled water, share in adecuated receptacles (tubes, bottles) and autoclave 5 minutes to 116°C before inoculate samples an their decimal dilutions. Shake to homogenize. Incubate 8-48 hours to 35-37°C aprox. (Total coliform and E.Coli) or to 44°C aprox. (Faecal coliform and E. Coli). Inoculated samples serve as well as transport medium, so it is not necessary incubate immediately after taking sample, and it can pass some hours between taking and incubation. Turn tu blue colour (Xgal): Presence of Coliforms (total or faecal according temperature of incubation). Blue Fluorescence (blue light by MUG) which can be observed in darkness with a U.V.A. 366 nm (lamp VMT050): E.Coli (confirm with 0.5 ml of KOVACS (SDA056), red ring of indol in surface is +). E.Coli O157 H7 is detected because is Xgal + (turns to blue), MUG – (no fluorescence) and indol + (red ring).

Bottles P/A with MCC:
Left: coliforms positive,
Right: coliforms Negative



Final user is the only responsible of destruction of grown microorganism according the current environmental legislation. Autoclave before throw to the rubbish.

Last revision made on May 2010